

Datasheet for R401-0050**Chicken RBCs 10% Washed Pooled Cells****Overview**

Description:	Chicken Red Blood Cells (RBC) 10% Washed Pooled Cells - R401-0050
Item No.:	R401-0050
Size:	50 mL
Applications:	ChIP, Other
Origin:	Chicken

Product Details

Background:	Chicken whole blood is washed to remove the platelet rich plasma, buffy coat layer, and leukocytes (WBC). Red blood cells are supplied as a 10 percent suspension in phosphate buffered saline (PBS). Chicken red blood cells are useful for the titration of complement, adsorption procedures, testing for agglutinins/HA assays, and for the preparation of stroma as particulate reagents. Chicken red blood cells are perishable and are collected and processed upon receipt of your order.
Synonyms:	Chicken Washed Pooled Cells, Chicken WPCs, Chicken Red Blood Cells, Chicken RBCs, erythrocytes
Species of Origin:	Chicken

Target Details

Relevant Links:	SDS
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Application Details

Suggested Applications:	ChIP, Other (Based on references)
Application Note:	Chicken whole blood cells are used for complement titration, adsorption procedures, HA assays and for the preparation of stroma as particulate reagents.
Assay Dilutions:	All assays should be optimized by the user. Recommended dilutions (if any) may be listed below.

Tissue Data

Tissue Type:	Red Blood Cells
Sex:	Mixed
Strain:	Chicken - Mixed

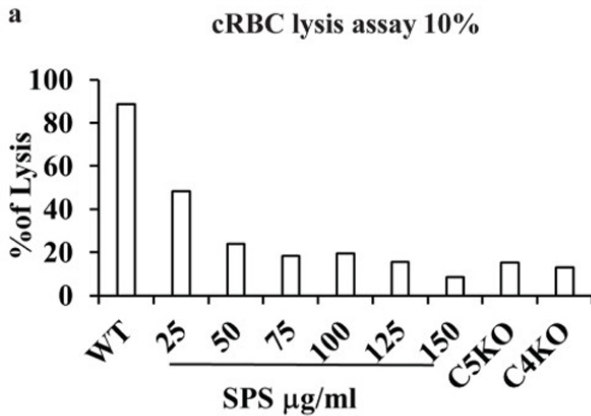
Formulation

Physical State:	Liquid
Buffer:	0.02 M Potassium Phosphate, 0.15 M Sodium Chloride, pH 7.2
Sterility:	Non-sterile
Preservative:	None
Stabilizer:	None

Shipping & Handling

Shipping Condition:	Wet Ice
Storage Condition:	Store chicken washed pooled red blood cells at 4° C prior to opening. Be advised that blood is a perishable product and exact shelf may depend on application.
Expiration:	This product MAY be stable for up to two (2) weeks if properly stored and handled.

Images

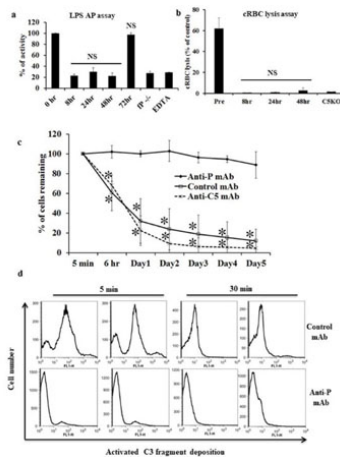


Agglutination

Assessment of lectin and classical pathway complement activity in Masp3^{-/-} mouse plasma.

(a) Sodium polyanethole sulfonate (SPS) dose-dependently inhibited mouse classical pathway complement activity in a chicken red blood cell (cRBC) hemolytic assay. Percentage of cRBC lysis was normalized to a cRBC sample completely lysed by hypotonic shock in double-distilled water. Mouse serum was diluted to 10% and incubated with Ab-sensitized chicken RBCs (p/n R401-0050) at 37°C for 30 min. Ab-sensitized chicken RBCs were prepared by incubating the cells with rabbit anti-chicken RBC Ab (p/n 103-4139) at 150 $\mu\text{g/ml}$ on ice for 1 h.

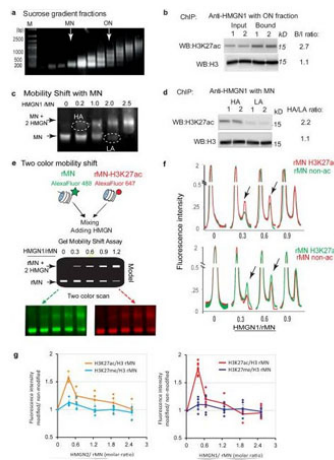
Fig 2. PMID: 36988282.



Figure

Therapeutic efficacy of anti-human P mAb 19.1 in a murine model of extravascular hemolysis: Pharmacodynamics of mAb 19.1 in hP transgenic mice. Each mouse was treated with 0.5 mg mAb 19.1 (n=3 mice). Serum samples were collected before and at various time points after mAb treatment and assessed for LPS-dependent AP complement activity. At this dosage of mAb 19.1, AP complement activity was suppressed to background (P^{-/-}) level for 2 days. EDTA: time 0 sample with EDTA (20 mM) added. NS, non-significant comparing 8, 24 and 48 hr samples with fP^{-/-} or EDTA-treated serum, or comparing 72 hr sample with 0 hr sample. One-way ANOVA. b: Pharmacodynamics of anti-mouse C5 mAb (BB5.1) in hP transgenic mice. Each mouse was treated with 1mg of anti-mouse C5 mAb (n=3). Serum samples were collected before and at various time points after mAb treatment and assessed for lytic activity using antibody-sensitized chicken RBCs. C5 knockout (KO) mouse serum was used as a control for C5 inhibition. Percentage of chicken RBC (cRBC) lysis was normalized to a sample completely lysed by hypotonic shock in double distilled water. * p<0.0001, NS: non-significant compared with C5KO serum. One-way ANOVA. c. Effect of anti-hP mAb 19.1 on the survival of transfused CFSC-labeled DAF^{-/-}/Crry^{-/-}/C3^{-/-} mouse erythrocytes in hP- transgenic mice. Recipient mice were treated with anti-hP mAb 19.1 (n=4) or an isotype control mAb (n=4) or anti-mouse C5 mAb (n=3) 6 hours prior to red blood cell transfusion and blood samples were taken at 5 min, 6 hrs and then daily for 5 days. The percentage of CFSC-labeled red blood cells was measured by FACS and normalized to that determined at 5 min (100%). Transfused

DAF^{-/-}/Crry^{-/-}/C3^{-/-} mouse red blood cells were rapidly eliminated in control mAb- or anti-C5 mAb-treated hP transgenic mice but such an outcome was prevented by mAb 19.1 treatment. * $p < 0.001$, Two-way ANOVA. d. FACS analysis of activated C3 fragment deposition on DAF^{-/-}/Crry^{-/-}/C3^{-/-} RBCs 5 and 30 min after their transfusion into control mAb- or mAb 19.1-treated hP-Tg/P^{-/-} mice (representative data from two recipient mice are shown). At both time points, C3 fragment deposition was significantly higher on RBCs transfused into control mAb-treated than mAb 19.1-treated hP-Tg/P^{-/-} mice. The reason for the marked reduction in C3 fragment deposition on RBCs in control mAb-treated hP-Tg/P^{-/-} mice between 5 and 30 min is unknown, but could be caused by C3 fragment degradation to C3d or shedding from the cell surface or rapid removal of the C3-opsonized cells. Data in a-c are presented as mean (SD) with indicated n numbers. Antibody-sensitized chicken RBC (p/n R401-0050) prepared by incubating the cells with a rabbit anti-chicken RBC antibody (p/n 103-4139). Fig 4. PMID: 29898960.



Figure

Preferential binding of HMGN1 to chromatin particles containing H3K27ac.

a Agarose gel showing sucrose gradient fractionated salt stripped MEF chromatin particles. MN: mononucleosomes, ON: Oligonucleosome (mostly tri-penta nucleosomes). b Western analysis of total ON (Input) and HMGN1 immunoprecipitated ON (bound) (c) Gel mobility shift -assay. Purified HMGN1 was added to salt stripped chicken erythrocyte MNs at the ratio indicated on top of each column. The MNs shifted at low HMGN1:MN were designated as high affinity (HA) while the MNs not shifted at high HMGN:MN were designated as low affinity (LA). d Western analysis of HA and LA mononucleosomes (MN). e Two color gel mobility shift assays of recombinant mononucleosomes (rMN). A mix of equal amounts of fluorescently Alexa 488 labeled rMN (green) and Alexa 647 labeled rMNH3K27ac (red) were incubated with various amounts of HMGN1, the mixture fractionated on native polyacrylamide gels, and the gels scanned to visualize and quantify either the red or green fluorescence. Shown is the experimental design and gel images visualized with red or green channels (f) Scan of the gels shown in e and of a similar gel in which the fluorescent labels are reversed. Top: Alexa 647 labeled rMNH3K27ac (red) and Alexa 488 labeled rMN (green). Bottom: Alexa 488 labeled rMNH3K27ac and Alexa 647 labeled rMN. Arrows point to preferential binding of HMGN1 to rMNH3K27ac in the shifted nucleosome. (g) Quantification of the scans shown in panel F and in Supplementary Fig. 4 for HMGN1 and of similar experiments done with HMGN2. Note that at low ratio of HMGN to nucleosomes, both HMGN1 and HMGN2 preferentially bind to rMNH3K27ac but not to rMNH3K27me3. Error bars represent corrected standard deviation, $n \geq 3$. Chicken Red Blood Cells (RBC) 10% Washed Pooled Cells (p/n R401-0050). Fig 2. PMID: 35197580.

References

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