

## Datasheet for 600-106-215

**GFP (GOAT) Antibody Biotin Conjugated****Overview**

|                      |  |
|----------------------|--|
| <b>Description:</b>  | Anti-GFP (GOAT) Antibody Biotin Conjugated - 600-106-215       |
| <b>Item No.:</b>     | 600-106-215  |
| <b>Size:</b>         | 1 mg   |
| <b>Applications:</b> | ELISA, WB, EM, FC, IF, IHC, IP, Multiplex, Other, Purification |
| <b>Reactivity:</b>   | GFP, eGFP, rGFP  |
| <b>Host Species:</b> | Goat   |

**Product Details**

|                      |   |
|----------------------|---|
| <b>Background:</b>   | Conjugated Anti-GFP is ideal for western blotting, ELISA and Immunohistochemistry. Green fluorescent protein is a 27 kDa protein produced from the jellyfish <i>Aequorea victoria</i> , which emits green light (emission peak at a wavelength of 509nm) when excited by blue light. GFP is an important tool in cell biology research. GFP is widely used enabling researchers to visualize and localize GFP-tagged proteins within living cells without the need for chemical staining. |
| <b>Synonyms:</b>     | goat anti-GFP Antibody biotin Conjugation, biotin conjugated goat anti-GFP antibody, Green Fluorescent Protein, GFP antibody, Green Fluorescent Protein antibody, EGFP, enhanced Green Fluorescent Protein, <i>Aequorea victoria</i> , Jellyfish  |
| <b>Host Species:</b> | Goat  |
| <b>Conjugate:</b>    | Biotin  |
| <b>Clonality:</b>    | Polyclonal  |
| <b>Format:</b>       | IgG   |

**Target Details**

|                        |   |
|------------------------|---|
| <b>Reactivity:</b>     | GFP, eGFP, rGFP   |
| <b>Immunogen Type:</b> | Recombinant Protein   |
| <b>Immunogen:</b>      | Recombinant Green Fluorescent Protein (GFP) fusion protein corresponding to the full length amino acid sequence (246aa) derived from the jellyfish <i>Aequorea victoria</i> . |

**Purity/Specificity:** Anti-GFP was prepared from monospecific antiserum by immunoaffinity chromatography using Green Fluorescent Protein (*Aequorea victoria*) coupled to agarose beads followed by solid phase adsorption(s) to remove any unwanted reactivities. Assay by immunoelectrophoresis resulted in a single precipitin arc against anti-Goat Serum, anti-biotin and purified and partially purified Green Fluorescent Protein (*Aequorea victoria*) Serum. No reaction was observed against Human, Mouse and Rat Serum Proteins.

**Relevant Links:**

- [UniProtKB - P42212](#)

## Application Details

|                                |  |
|--------------------------------|--|
| <b>Tested Applications:</b>    | ELISA, WB  |
| <b>Suggested Applications:</b> | EM, FC, IF, IHC, IP, Multiplex, Other, Purification (Based on references)  |
| <b>Application Note:</b>       | Anti-GFP Biotin Conjugated Antibody has been tested by ELISA and western blot and is suitable for immunoblotting, ELISA, immunohistochemistry, immunomicroscopy as well as other antibody based assays using streptavidin or avidin conjugates requiring lot-to-lot consistency. |
| <b>Assay Dilutions:</b>        | All assays should be optimized by the user. Recommended dilutions (if any) may be listed below.  |
| <b>ELISA:</b>                  | 1:50,000 - 1:80,000  |
| <b>IHC:</b>                    | 1:1,000 - 1:5,000  |
| <b>WB:</b>                     | 1:2,000 - 1:10,000   |

## Formulation

|                               |  |
|-------------------------------|--|
| <b>Physical State:</b>        | Lyophilized  |
| <b>Concentration:</b>         | 1.0 mg/mL by UV absorbance at 280 nm                                   |
| <b>Buffer:</b>                | 0.02 M Potassium Phosphate, 0.15 M Sodium Chloride, pH 7.2             |
| <b>Preservative:</b>          | 0.01% (w/v) Sodium Azide   |
| <b>Stabilizer:</b>            | 10 mg/mL Bovine Serum Albumin (BSA) - Immunoglobulin and Protease free |
| <b>Reconstitution Volume:</b> | 1.0 mL   |
| <b>Reconstitution Buffer:</b> | Restore with deionized water (or equivalent)                           |

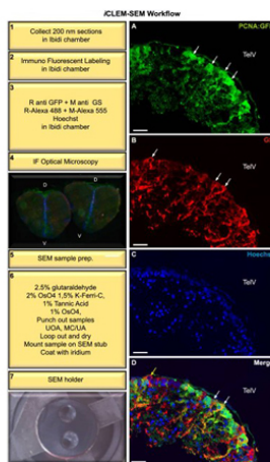
## Shipping & Handling

**Shipping Condition:** Ambient

**Storage Condition:** Store Anti-GFP at 4° C prior to restoration. For extended storage aliquot contents and freeze at -20° C or below. Avoid cycles of freezing and thawing. Centrifuge product if not completely clear after standing at room temperature. This product is stable for several weeks at 4° C as an undiluted liquid. Dilute only prior to immediate use.

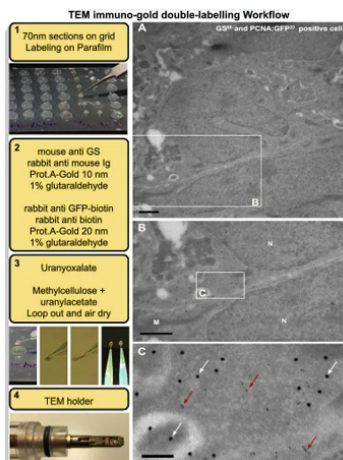
**Expiration:** Expiration date is one (1) year from date of receipt.

## Images



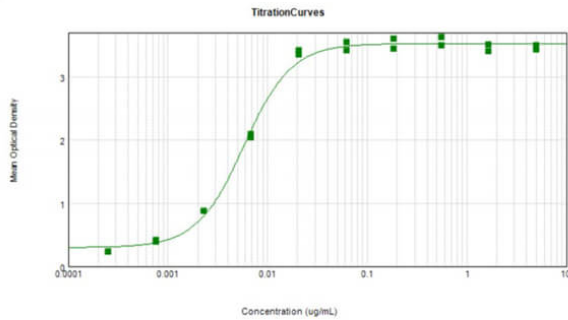
### Immunofluorescence Microscopy

iCLEM-SEM workflow and immunofluorescent labelling on Tokuyasu chemical-fixed cryosections. Left: Major steps required for iCLEM-SEM workflow (steps 1–6). (A) Green Fluorescent Protein (GFP)-positive labelling (green) of proliferating cells from the Tg(PCNA:GFP) transgenic line (white arrows). (B) Glutamine Synthetase (GS)-positive labelling (red) of glial cells (white arrows). (C) Hoechst counterstaining (blue). (D) Multi-channel merge showing co-labelling of PCNA:GFP-positive cells with the glial marker (orange cells; white arrows) in the cytoplasm of a small number of cells. Differences in the intensity of antibody staining can additionally be noted (compare white arrows and yellow arrow). D dorsal; V ventral. TelV telencephalic ventricle. Scale bar (A–D) 20 μm. Fig 2. PMID: 33441723



### Immunofluorescence Microscopy

TEM immuno-gold double-labelling workflow and example of positive labelling on Tokuyasu chemical-fixed cryosections. Left: Major steps (1–4) required for TEM immuno-gold double-labelling workflow and associated images. (A,B,C) Cytosolic PCNA:GFP+/GS+ immuno-gold double-labelling at low (A) and high (B,C) magnifications demonstrating co-localization of the 10 nm (GS; red arrowheads) and 20 nm (PCNA:GFP; white arrowheads) particles. N, nucleus. Scale bar (A) 2 μm; (B,C) 200 nm. Fig 6. PMID: 33441723



### ELISA

ELISA Results of Goat Anti-GFP Antibody tested against purified GFP protein. Each well was coated in duplicate with 10µg/mL of GFP protein (p/n 000-001-215). The starting dilution of antibody was 5µg/mL and the X-axis represents the Log10 of a 3-fold dilution. This titration is a 4-parameter curve fit where the IC50 is defined as the titer of the antibody. Assay performed using 3% Fish Gel/PBS Blocking buffer (p/n MB-066) and SA-HRP conjugated (p/n S000-03).

## References

- Ismaeel A et al. Division-Independent Differentiation of Muscle Stem Cells During a Growth Stimulus. *Stem Cells*. (2024)
- Daw TB et al. Direct Comparison of Epifluorescence and Immunostaining for Assessing Viral Mediated Gene Expression in the Primate Brain. *Hum Gene Ther*. (2023)
- Lu H et al. Alternative splicing and heparan sulfation converge on neurexin-1 to control glutamatergic transmission and autism-related behaviors. *Cell Rep*. (2023)
- Kim YE et al. Reversibility and developmental neuropathology of linear nevus sebaceous syndrome caused by dysregulation of the RAS pathway. *Cell Rep*. (2023)
- Stajano D et al. Tetraspanin  $\alpha 6$  depletion impairs extracellular vesicle docking at target neurons. *ISEV journals* (2023)
- De Mazière A et al. An optimized protocol for immuno-electron microscopy of endogenous LC3. *Autophagy*. (2022)
- Jones, T et al. Differential requirements for the Eps15 homology domain proteins EHD4 and EHD2 in the regulation of mammalian ciliogenesis. *Traffic (Copenhagen, Denmark)* (2022)
- Tran NH et al. The stress-sensing domain of activated IRE1 $\alpha$  forms helical filaments in narrow ER membrane tubes. *Science*. (2021)
- Oorschot V et al. TEM, SEM, and STEM-based immuno-CLEM workflows offer complementary advantages. *Sci Rep*. (2021)
- Zulkefli K et al. A role for Rab30 in retrograde trafficking and maintenance of endosome-TGN organization. *Exp Cell Res*. (2021)
- Wieghofer P et al. Mapping the origin and fate of myeloid cells in distinct compartments of the eye by single-cell profiling. *EMBO J*. (2021)
- Li H et al. Cellular requirements for PIN polar cargo clustering in *Arabidopsis thaliana*. *New Phytol*. (2021)
- Bohlen MO et al. Adeno-Associated Virus Capsid-Promoter Interactions in the Brain Translate from Rat to the Nonhuman Primate. *Hum Gene Ther*. (2020)

- Cabukusta B et al. Human VAPome Analysis Reveals MOSPD1 and MOSPD3 as Membrane Contact Site Proteins Interacting with FFAT-Related FFNT Motifs. *Cell Rep.* (2020)
- Jongsma ML et al. SKIP-HOPS recruits TBC1D15 for a Rab7-to-Arl8b identity switch to control late endosome transport. *EMBO J.* (2020)
- Cushnie AK et al. Using rAAV2-retro in rhesus macaques: promise and caveats for circuit manipulation. *J Neurosci Methods.* (2020)
- Bohlen MO, El-Nahal HG, Sommer MA. Transduction of Craniofacial Motoneurons Following Intramuscular Injections of Canine Adenovirus Type-2 (CAV-2) in Rhesus Macaques. *Front Neuroanat.* (2019)
- Andres-Alonso M, Ammar MR, Butnaru I, et al. SIPA1L2 controls trafficking and local signaling of TrkB-containing amphisomes at presynaptic terminals. *Nat Commun.* (2019)
- Croop B et al. Facile single-molecule pull-down assay for analysis of endogenous proteins. *Phys Biol.* (2019)
- Yousefi OS et al. Optogenetic control shows that kinetic proofreading regulates the activity of the T cell receptor. *Elife.* (2019)
- Lee JY et al. Limiting Neuronal Nogo Receptor 1 Signaling during Experimental Autoimmune Encephalomyelitis Preserves Axonal Transport and Abrogates Inflammatory Demyelination. *J Neurosci.* (2019)
- Jangphattananont N et al. Distinct localization of mature HGF from its precursor form in developing and repairing the stomach. *Int J Mol Sci.* (2019)
- Koerver L et al. The ubiquitin-conjugating enzyme UBE 2 QL 1 coordinates lysophagy in response to endolysosomal damage. *EMBO Rep.* (2019)
- Ge Y et al. Clptm1 limits forward trafficking of GABAA receptors to scale inhibitory synaptic strength. *Neuron.* (2018)
- McArthur K et al. BAK/BAX macropores facilitate mitochondrial herniation and mtDNA efflux during apoptosis. *Science.* (2018)
- Bender J et al. Multiplexed antibody detection from blood sera by immobilization of in vitro expressed antigens and label-free readout via imaging reflectometric interferometry (iRif). *Biosens Bioelectron.* (2018)
- Maeder CI, Kim JI, Liang X, et al. The THO Complex Coordinates Transcripts for Synapse Development and Dopamine Neuron Survival. *Cell.* (2018)
- Zöllner et al. Silencing of TGF $\beta$  signalling in microglia results in impaired homeostasis. *Nature Communications* (2018)
- Liebmann et al. Regulation of Neuronal Na,K-ATPase by Extracellular Scaffolding Proteins. *International Journal of Molecular Sciences* (2018)
- Bower NI et al. Mural lymphatic endothelial cells regulate meningeal angiogenesis in the zebrafish. *Nat Neurosci.* (2017)
- Goldmann et al. Origin, fate and dynamics of macrophages at central nervous system interfaces. *Nature Immunology* (2016)
- Sztal et al. Zebrafish models for nemaline myopathy reveal a spectrum of nemaline bodies contributing to reduced muscle function. *Acta Neuropathologica* (2015)
- Baek et al. An AKT3-FOXG1-reelin network underlies defective migration in human focal malformations of cortical development. *Nature Medicine* (2015)

- Henau et al. A redox signalling globin is essential for reproduction in *Caenorhabditis elegans*. *Nature Communications* (2015)
- Oorschot VMJ et al. Immuno correlative light and electron microscopy on Tokuyasu cryosections. *Methods Cell Biol.* (2014)
- Kang Y et al. A combined transgenic proteomic analysis and regulated trafficking of neuroligin-2. *J Biol Chem.* (2014)
- Goldmann T et al. A new type of microglia gene targeting shows TAK1 to be pivotal in CNS autoimmune inflammation. *Nat Neurosci.* (2013)
- Kunte, A et al. Endoplasmic reticulum glycoprotein quality control regulates CD1d assembly and CD1d-mediated antigen presentation. *The Journal of Biological Chemistry* (2013)
- Jain A et al. Single-molecule pull-down for studying protein interactions. *Nat Protoc.* (2012)
- Panter MS et al. Dynamics of major histocompatibility complex class I association with the human peptide-loading complex. *J Biol Chem.* (2012)
- Jain, A et al. Probing cellular protein complexes using single-molecule pull-down. *Nature* (2011)
- Graf ER et al. Neurexins induce differentiation of GABA and glutamate postsynaptic specializations via neuroligins. *Cell.* (2004)

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